

Program & Abstracts

International Conference Food for a Quality Life

Jakarta - Indonesia
15 - 16 October 2014

Organized by:



SEAFAST
CENTER



Department of
Food Science
& Technology

In conjunction with:



Food ingredients
Asia

Supported by:



FISTA



Media partner:

FOODREVIEW
INDONESIA





SEAFAST
CENTER



Department of
Food Science
& Technology

International Conference
Food for a Quality Life
October 15 - 16, 2014, Jakarta - Indonesia

CONTENTS

| | |
|--|-----|
| INTRODUCTION | 1 |
| ORGANIZERS | 4 |
| PROGRAM..... | 7 |
| POSTER PRESENTATION | 19 |
| SPEAKER PROFILES..... | 26 |
| ABSTRACTS OF ORAL PRESENTATION | |
| Keynote Speech : The Role of Food for a Quality Life | 39 |
| Plenary. | 40 |
| New Challenges in Food Industry | 55 |
| Innovations in Managing Food Safety and Quality..... | 73 |
| Advances in Nutrition and Health..... | 81 |
| Emerging Food Issues | 97 |
| ABSTRACTS OF POSTER PRESENTATION | |
| Emerging Food Issues | 101 |
| New Challenges in Food Industry | 103 |
| Innovations in Managing Food Safety and Quality..... | 124 |
| Advances in Nutrition and Health..... | 132 |
| ABSTRACTS OF SPECIAL EVENT | |
| Graduate Students Research Paper Competition..... | 143 |
| PARTICIPANTS..... | 149 |

APPLICATION OF BACTERIOPHAGES COCKTAIL TO CONTROL *Salmonella typhimurium* IN CHICKEN MEAT

Agustin Krisna Wardani*, Aji Sutrisno, Alfi Azizah, Zein Nurriskiawan
Jurusan Teknologi Hasil Pertanian, Fakultas Teknologi Pertanian,
Universitas Brawijaya
Jl. Veteran Malang 65145
Email: agustinwardani@ub.ac.id

ABSTRACT

Food safety is an increasingly important public health issue. Many efforts are intensified all over the world to improve food safety. The number of cases of several foodborne diseases has been rising continuously in many countries. Salmonellosis is one of the most common and widely distributed foodborne diseases and is caused by *Salmonella*. Antibiotics are usually used to tackle the problem of foodborne diseases. However, antibiotic have adverse effects such as bacterial resistance to antibiotics as well as killing the entire colony of microbes in the gut both beneficial and adverse. The use of lytic bacteriophages for the bio-control of foodborne pathogens should be a promising strategy to overcome this problem. In this study, the effectiveness of a lytic bacteriophage cocktail ($\Phi 14$) isolated from chicken intestine was determined in chicken meat experimentally contaminated with *Salmonella typhimurium*.

Production of $\Phi 14$ with the concentration of $8,3 \times 10^3$ PFU/ml was conducted by using *Shigella flexneri* as a host. To obtain the effective application of bacteriophage, the optimization of host infection was performed. A 30 minutes infection of bacteriophage to cell target (*S. typhimurium*) was found to be an optimum time to lyze the cell target. A reduction of *S. typhimurium* (1-3 log) was obtained after infection of bacteriophage in chicken meat under 37°C for 2, 4, 6 days. This result shows that the potential effectiveness of this bacteriophage cocktail as a biocontrol agent of *S. typhimurium* in food product under conditions similar to those used in their production.

Keywords: foodborne diseases, bacteriophage, bio-control, *Salmonella typhimurium*, chicken meat

INTRODUCTION

Food safety issues have become a global problem, so it received great attention in the establishment of public health policy. Diseases caused by consuming foods that contain pathogenic bacteria (foodborne disease) and incidence of food contamination events occur not only in developing countries where sanitation and hygiene conditions are generally poor but also in developed countries. An estimated one in three people resident in developed countries experienced food poisoning each year. (Secretariat General of the Food Intelligence Network, 2005). During 2004, based on reports Center / POM throughout Indonesia have occurred an outbreak (KLB), food poisoning as many as 153 events in 25 provinces (Secretariat General of the Food Intelligence Network, 2005). In Indonesia from 18 cases of food poisoning that occurred in 2003, 83.30% are caused by pathogenic bacteria.

Salmonella is a bacteria that can cause typhoid, paratyphoid, and salmonellosis. In Europe and the United States cases of illness caused by Salmonella enteritidis is transmitted through chicken meat, eggs, and dairy products (Baumler *et al.*, 2000). Salmonella are bacteria that cause intestinal infections second most common in the United States. More than 7,000 cases of Salmonellosis were confirmed in 2009, but most cases are not reported (Djafaar and Rahayu, 2004). To overcome the problem of foodborne disease are usually people use antibiotics. But antibiotics have been used extensively by people had detrimental effects such as bacterial resistance to antibiotics as well as the killing of entire colonies of microbes in the gut both beneficial and adverse. As an alternative to antibiotics it is used bacteriophages. Bacteriophage has the advantage of its ability to specifically attack bacteria with high specificity without disturbing other microbiota. Bacteriophages are highly specific nature is beneficial because it makes safe when consumed by humans. Based on the United States Food and Drug Administration (2006) bacteriophages on cheese can be used to kill *Listeria monocytogenes* and classified in the GRAS (Generally Recognized As Safe) and can be used in food products.

Meat products, including chicken meat is a food product susceptible to pathogen contamination, one Salmonella typhimurium. Based on the advantages offered by bacteriophages so in the present study using bacteriophage applied to chicken meat after contaminated with *Salmonella typhimurium*. The aim of this study was to determine the optimum time of bacteriophages in lysing *Salmonella typhimurium* in chicken meat as well as knowing the amount of reduction in *Salmonella typhimurium*.

MATERIALS AND METHOD

Materials

Samples used are fresh chicken meats, bacteriophages $\Phi 14$, *Salmonella typhimurium*, *Salmonella flexneri* as a host. The chemicals used are CaCl_2 , NaCl , distilled water, alcohol, methylated spirits, PP plastic, aluminum foil, cotton.

Bacteriophage production

Seven ml Nutrient Broth inoculated with 200 μL of bacterial host then incubated for 90 minutes under 37°C . After it was added 40 μL and 250 μL CaCl_2 bacteriophages, incubated for four to six hours, then observed the level of turbidity. Then the liquid is centrifuged at 6000 rpm, 5°C for 10 minutes. The filtrate formed was filtered with a sterile filter of 0.2 μm .

Optimization of *Salmonella typhimurium* infection

75 ml of sterile Nutrient Broth were inoculated with 200 μl of pathogenic bacteria and then incubated for 0, 30, 60 and 90 minutes. After that 100 μL CaCl_2 , 200 μL bacteriophage $\Phi 14$ were added. The absorbance was measured every 2 hours for the control (pathogens without infection of bacteriophage $\Phi 14$) and pathogens with infection of bacteriophage $\Phi 14$.

Application of bacteriophage in meat

Fresh meat was sterilized at 121°C for 15 minutes, then meat was homogenized by using stomacher for 15 minutes. Fifteen ml of aliquots were taken and added with 50 μl of *Salmonella typhimurium*, incubated at 37°C for 0,30,60 minutes. One hundreds (100 μl) of 0.3 M of CaCl_2 and 500 μl of bacteriophages $\Phi 14$ were further added then incubated at 37°C incubated for 0,2,4,6 days. Aliquots were diluted to a certain dilution. Then grown in SSA media using spread plate method, under 37°C for 24 hours. The number of bacteria were counted in CFU / ml.

Data analysis

The data obtained were analyzed using descriptive analysis method to investigate the addition of a bacteriophage (30, 60, and 90 minutes) to lyse the growth of *Salmonella typhimurium*. The lysis can be observed by comparing the growth of *Salmonella* with phage infection and without phage infection.

RESULT AND DISCUSSION

Bacteriophage production

Bacteriophages $\Phi 14$ were used in this study were obtained from the isolation in previous research (Zein, 2011). Isolation of bacteriophages is intended to obtain mixture of virulent bacteriophages (phage cocktail) capable of inhibiting the growth of pathogenic bacteria. Bacteriophages isolated from the intestinal samples of fresh chicken, ground, salted vegetables, and garbage. Isolation of bacteriophages and host was done with double layer method. From the results showed that one isolate as a host number 14 was successfully isolated from the intestinal samples (U14). Bacteriophage production is done to obtain high concentration of bacteriophage which is enough to be used for application in chicken meat. Production was done by inoculating host (*Shigella flexnery*) in 7 ml NB then incubation was performed at 37°C for 90 minutes to allow *S. flexnery* grow. Subsequently, it was added CaCl₂ and bacteriophages $\Phi 14$, and then incubated for 8 hours. The observation was done every half an hour until the lysis mechanism were observed. Lysis occurred when solution become clear, when compared to the control (without addition of bacteriophage). Table 1 show the degree of turbidity (lysis) of the solution.

Table 1. Turbidity of solution (lysis) during 8 hours incubation

| Time (min) | Control (without $\Phi 14$ infection) | Treated sample (with $\Phi 14$ infection) |
|------------|--|--|
| 0 | + | + |
| 30 | + | + |
| 60 | + | + |
| 90 | + | + |
| 120 | + | + |
| 150 | ++ | + |
| 180 | ++ | + |
| 210 | +++ | ++ |
| 240 | +++ | ++ |
| 270 | +++ | ++ |
| 300 | ++++ | ++ |
| 330 | ++++ | ++ |
| 360 | ++++ | ++ |
| 390 | ++++ | +++ |
| 420 | +++++ | +++ |
| 450 | +++++ | ++++ |
| 480 | +++++ | ++++ |

Nb: + very clear; ++ clear; +++ slightly turbid; ++++ turbid; +++++ very turbid

Based on the result, it shows that the solution with bacteriophage $\Phi 14$ infection has more clearer than the control solution (without bacteriophage $\Phi 14$ infection) during 6 hours incubation. The level of solution clarity dictated the host lysis due to bacteriophage infection. To produce the phage, the liquid was centrifuged and filtered with 0.2 μm sterile filter to obtain lysate for a new stock. The lysis phenomenon can be observed visually as shown in figure 1.

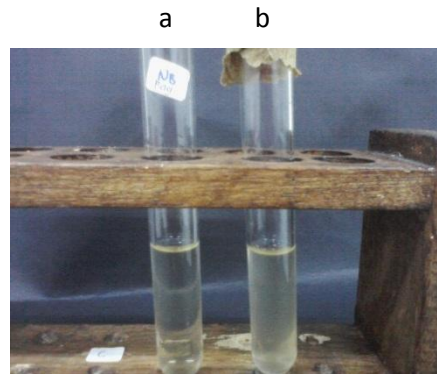


Figure 1.. Bacteriophage production. Solution with $\Phi 14$ infection (a); solution without $\Phi 14$ infection (b)

To determine the concentration of bacteriophages, double layer method was done (Chow *et al.* , 1988) . 15 ml of media hard agar NA (1.5 %) is poured dish and allowed to solidify. Then 3 ml of soft agar NA (0.7 %) was added 100 μL of 20 hours *Shigella flexneri*, 100 μL of 0.3 M CaCl_2 , and 100 μL of bacteriophages and then poured on top of the hard agar.

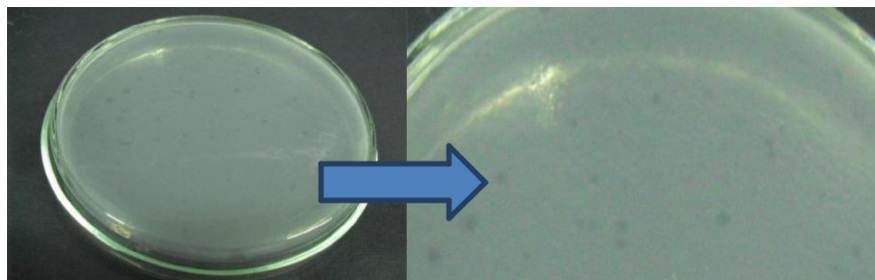


Figure 2. Plaque formation

Double layer was incubated at 37°C for 6 hours and observed the formation of plaque (clear zone on host bacteria). The formation of plaque is presented in Figure 2. Plaque is formed then calculated by unit PFU / ml (plaque forming units / ml) as many as 8.3×10^7 PFU / ml .

Optimization of *Salmonella typhimurium* lysis

The potency of bacteriophage $\Phi 14$ as antimicrobial should be tested against pathogens, *Salmonella typhimurium*. Before the application of bacteriophages $\Phi 14$ into chicken meat, it is important to determine the optimum time of bacteriophage infection in inhibiting *Salmonella typhimurium*. The measurement of absorbance values of *Salmonella* growth show the infection of bacteriophages against *S. typhimurium* (Table 2).

Table 2. Growth of *Salmonella typhimurium*

| Time (min) | OD 600 nm | OD 600 nm | OD 600 nm | OD 600 nm |
|------------|-----------------------------------|-----------------------------------|-----------------------------------|-----------------------------------|
| | Control (without phage infection) | With phage infection after 30 min | With phage infection after 60 min | With phage infection after 90 min |
| 0 | 0,024 | 0,029 | 0,0301 | 0 033 |
| 30 | 0,054 | 0,049 | 0,047 | 0,055 |
| 60 | 0,072 | 0,079 | 0,076 | 0,081 |
| 90 | 0,116 | 0,093 | 0,089 | 0,097 |
| 120 | 0,127 | 0,117 | 0,101 | 0,112 |
| 150 | 0,135 | 0,121 | 0,111 | 0,116 |
| 180 | 0,171 | 0,154 | 0,136 | 0,142 |
| 210 | 0,196 | 0,068 | 0,145 | 0,162 |
| 240 | 0,216 | 0,102 | 0,129 | 0,179 |
| 270 | 0,256 | 0,147 | 0,137 | 0,191 |

Table 2 shows that the bacteriophage $\Phi 14$ is able to inhibit *S. typhimurium*. Inhibition of bacteriophage is shown by the decline of the absorbance of *S. typhimurium* growth. This decrease indicates the lysis of *S. typhimurium*. Lysis of bacteria due to the bacteria receptors is compatible with bacteriophage receptors. This allowed the inserting of genetic material of bacteriophage into the bacteria and use it for bacteriophage multiplication. Bacteriophages can only enter the bacterial cell membrane receptors when the natural bacterial cell is suitable with bacteriophage receptor (Mayer, 2010). The inhibition of bacteriophage $\Phi 14$ to *S. typhimurium* is presented in figure 3.

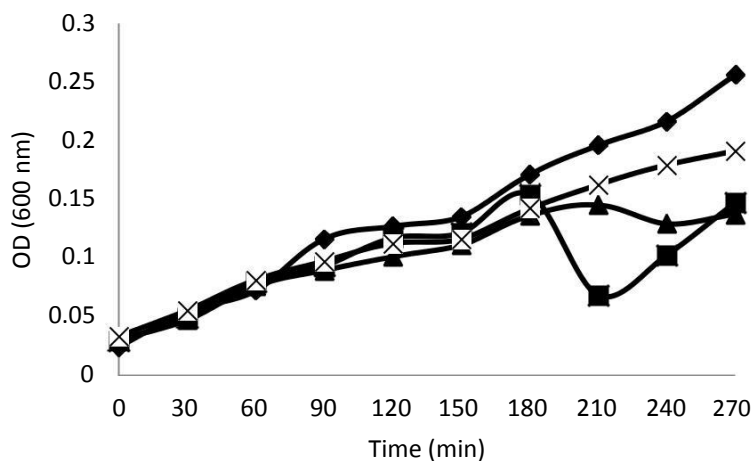


Figure 3. Inhibition of bacteriophage $\Phi 14$ to *Salmonella typhimurium*

- ◆ control (without $\Phi 14$ infection)
- $\Phi 14$ infection after 30 min; ▲ $\Phi 14$ infection after 60 min
- × $\Phi 14$ infection after 90 min

The variation of phage injection after 30 minutes, 60 minutes, and 90 minutes incubation of *S. typhimurium* is to give the time to grow of *S. typhimurium* which will be the optimum time of bacteriophage $\Phi 14$ in lysing *S. typhimurium*. Figure 3 shows that the OD values of control (without phage injection) have continued to rise indicating *S. typhimurium* has grown and multiply. In the case of the phage addition after 30 minutes incubation, it shows that OD value continues to rise from minute 0 to minute 180, then declined sharply until 210 minutes. This decrease indicates that there is a lot of *S. typhimurium* lysis of the bacteriophage $\Phi 14$ attack. During phage addition after 60 minutes incubation, the OD values have continued to rise and begins to decrease in minute 240, however not so great decrease was observed. At the time of the addition of phage after 90 minutes incubation, the OD value continues to increase but the increase is lower than control. This result show that the addition of phage 30 minutes is the best time for $\Phi 14$ to lyse *S. typhimurium* because the OD value was decline significantly, which means that most *S. typhimurium* lysis. According Todar (2008), the lysis of bacteria can occur every 15 minutes up to several days depending on the type of bacteria. From a bacteriophage that infects bacteria it will produce new phages 200-300 with effect become infected bacteria lyse (Ellis, 2007).

APPLICATION OF BACTERIOPHAGE IN CHICKEN MEAT

Bacteriophage $\Phi 14$ was applied to chicken meat to investigate the effectiveness of bacteriophage to inhibit or lyse *S. typhimurium*. According to Guenther et al., (2008) bacteriophage is a natural enemy of bacteria, have specific properties invade their host bacteria. Bacteriophage specific targets to eliminate

bacteria without interfering with other bacterial viability. It is desirable from an antimicrobial agent for use in food, because it can maintain the quality of products. In this study, a comparison of the growth of *S. typhimurium* on chicken meat without addition of bacteriophage and the growth of *S. typhimurium* on chicken meat with the addition of bacteriophages. In this study, the concentration of *S. typhimurium* which is applied in chicken meat was determined before the application of bacteriophages $\Phi 14$. Total number of *S. typhimurium* was calculated in SSA media using spread plate method, and the results obtained *S. typhimurium* concentration of $5,7 \times 10^4$ CFU / ml. The next step of the application, chicken meat was sterilized at 121°C for 15 minutes, to make chicken meat sterile from contaminant so that only *S. typhimurium* that will grow. Furthermore, chicken meat was homogenized for 120 seconds and the resulting aliquots were transferred to 10 ml centrifuge tube. Aliquots were then inoculated with *S. typhimurium* ($5,7 \times 10^4$ CFU / ml) and incubated for 0, 30, 60 minutes to give the time for *S. typhimurium* growing. Furthermore, the addition of CaCl_2 and bacteriophage $\Phi 14$ ($8,3 \times 10^3$ PFU / ml) and incubated for 0, 2, 4, 6 days. The next step, to apply the diluted aliquot in media SSA (Salmonella Shigella Agar) using a spread plate method then incubated at 37°C for 24 hours. Then calculated the number of Salmonella typhimurium that grows in plate. Plating results of *S. typhimurium* in SSA media is presented in Figure 4



Figure 4. Plating of *Salmonella typhimurium* in SSA media

Salmonella typhimurium calculation was done at 0, 2, 4, and 6 days incubation. The result of the calculation of *S. typhimurium* during 6 days incubation can be seen in Table 3.

Table 3. Number of *Salmonella typhimurium* during 6 days incubation

| Sample | Number of bacteria (CFU/ml) | | | |
|----------------------------------|-----------------------------|-------------------|-------------------|----------------------|
| | 0 day | 2 days | 4 days | 6 days |
| Without phage infection | $4,2 \times 10^3$ | $5,2 \times 10^7$ | $3,6 \times 10^8$ | $6,1 \times 10^9$ |
| With phage infection (0 min) | $3,5 \times 10^3$ | $4,1 \times 10^4$ | $7,3 \times 10^5$ | $5,8 \times 10^7$ |
| Without phage infection | $9,2 \times 10^3$ | $4,6 \times 10^6$ | $5,3 \times 10^8$ | $8,5 \times 10^9$ |
| With phage infection (30 min) | $8,6 \times 10^3$ | 5×10^5 | $5,2 \times 10^5$ | $9,3 \times 10^6$ |
| Without phage infection | $4,9 \times 10^4$ | $6,7 \times 10^7$ | $7,9 \times 10^9$ | $3,1 \times 10^{10}$ |
| With phage infection (60 min) | $3,7 \times 10^4$ | $4,1 \times 10^5$ | $5,3 \times 10^7$ | $5,6 \times 10^8$ |

Based on Table 5 the number of *S. typhimurium* were grown on chicken meat increased after for 6 days incubation, but the increase was reduced by the addition of bacteriophage $\Phi 14$. This is because the $\Phi 14$ is capable to lyse or kill *S. typhimurium* during incubation. On 2 day incubation, after addition of $\Phi 14$ (30 min), the number of *S. typhimurium* fell as much as 1.01 log, whereas the addition of phage (60 min) decreased as much as 2.21 log. Whereas, the addition of phage (0 min) shows the decrease of *S. typhimurium* reached at 3.10 log. This is because most of *S. typhimurium* has been attacked by bacteriophages $\Phi 14$. On 4 days incubation, the number of *S. typhimurium* after addition of phage (30 min) the decreased 3.01 log was observed, on the addition of phage 60 minutes the decrease at 2.17 log, whereas the addition of phage 0 minutes the decrease at 2.70. On the 6 days incubation, the number of *S. typhimurium* decreased until 2.96 log, on the addition of phage 60 minutes decrease until 1.75 log, while the addition of phage 0 minutes fell as much as 2.03 log. The lysis of pathogenic bacteria caused by the appropriate receptor of pathogens with the receptor of bacteriophage. This resulted in the suitability of a bacteriophage capable of inserting its genetic material into the bacteria and use it for multiplication. According to Mayer (2010) Bacteriophages can only enter the bacterial cell membrane where the receptor naturally bacterial cells is consistent with the receptor of bacteriophage. In addition, the bacteriophage does not cause a change in the organoleptic (color, odor, flavor, and structure) of the food product, therefore bacteriophages can be used as a controller of harmful pathogenic bacteria in food product (Hagens, 2007). Lysis curve of *S. typhimurium* attacked by bacteriophage $\Phi 14$ in chicken meat during incubation is presented in Figure 5.

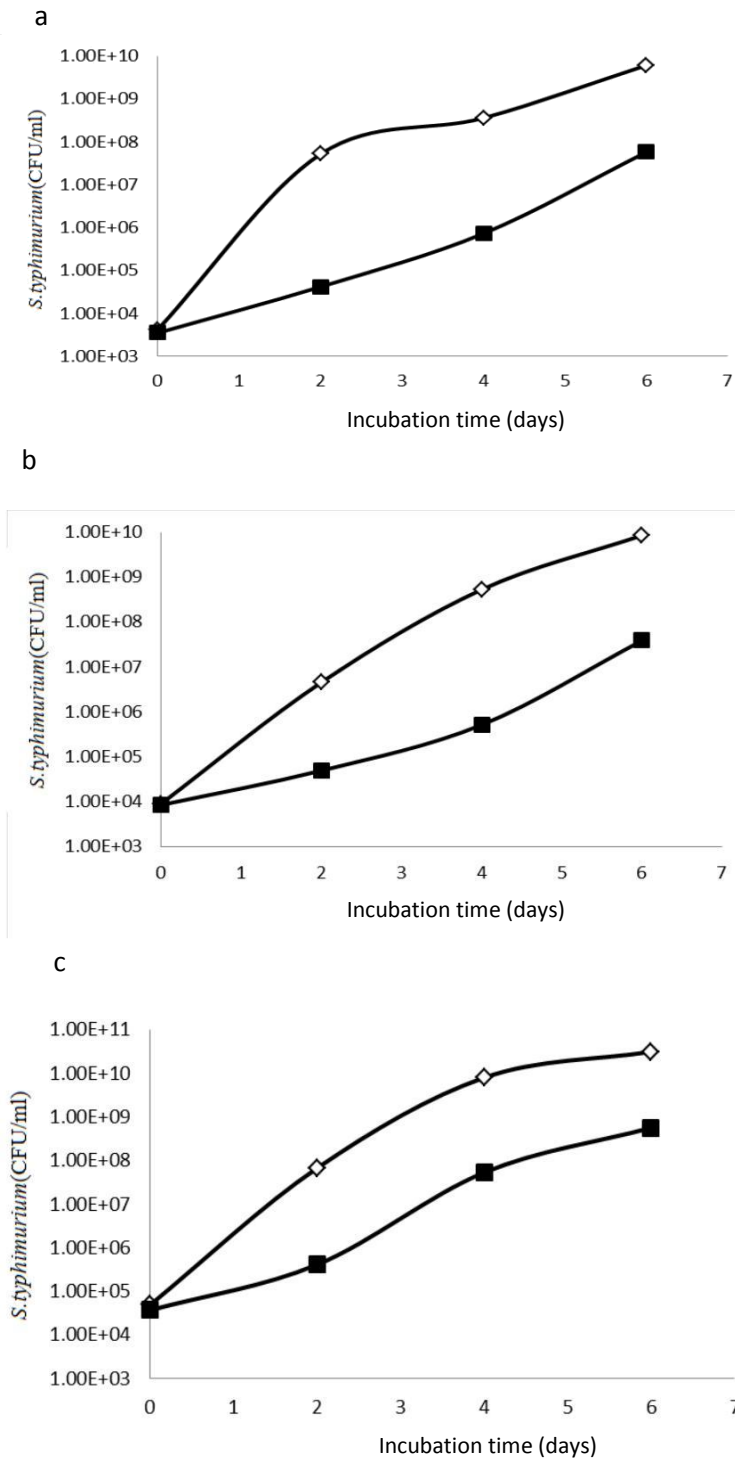


Figure 5. Lysis curve of *S. typhimurium* in chicken meat; phage addition at 0 minute (a); phage addition at 30 minutes (b); phage addition at 60 minutes (c). \diamond Control (without phage addition); \blacksquare (with phage addition)

Based on the pictures it show that the number of *S. typhimurium* in control (without addition of bacteriophage Φ 14) always higher than the number in sample with the addition of bacteriophage Φ 14.

Meaning that bacteriophage Φ 14 capable of inhibiting or reducing the growth of *S. typhimurium* during 6 days of incubation. After 6 days of incubation, the highest reduction of *S. typhimurium* was obtained with phage addition at 30 minutes. While on the 2nd day of incubation, the addition of phage at 0 minutes show the highest reduction of *S. typhimurium*. According to Leverentz *et al.*, (2004) a high concentration of bacteriophages are important in reducing the number of bacterial targets. In addition, the addition of bacteriophage also plays an important role in reducing the amount of bacteria during storage. In the study conducted by Guenther, *et al.* (2008) the application of bacteriophage A511 (3×10^8 PFU / ml) in milk chocolate that has been contaminated with *Listeria monocytogenes* (1×10^3 CFU / ml), with a time addition of bacteriophage at 1 hour, after 6 days of storage (20^0C) the amount of *Listeria monocytogenes* is able to be reduced as much as 6.4 log units.

CONCLUSION

Bacteriophages Φ 14 has the potency to be applied as bio-control for *Salmonella typhimurium* in meat product. It has the capacity to reduce the number of *S. typhimurium* as much as 2.96 log. The most optimum time of inhibition of bacteriophages against *Salmonella typhimurium* was observed at 30 minutes after incubation.

REFERENCES

- Ackermann, W. 2006. **Clasification of Bacteriophage 2nd edition**. Faculty of Medicine. Laval University. Quebec Canada. www.thebacteriophages.org. Diaskes 1 Februari 2012
- Anonymous^a. 2012. **Struktur Virus**. www.filzahazny.wordpress.com. Diaskes tanggal 1 februari 2012
- Anonymous^b. 2012. **Daur Hidup Bakteriofag**. www.ramsites.net. Diaskes 1 februari 2012
- Betsy, W. F. 2007. **Foodborne listeriosis**. Clin. Infect. Dis. 31:770-775
- Carlton, R. M., W. H. Noordman, B. Biswas, E. D. de Meester, and M. J. Lossner. 2005. **Bacteriophages P100 for Control of Listeria monocytogenes in Foods: genome sequence, bioinformatic analyses, oral toxicity study, and application**. Regul. Toxicol. Pharmacol. 43:301-312
- Coffey, J.J., C. A. Batt, and A. J. Sinskey. 1988. **Characterization of Bacteriophages chapter 3**. Appl. Environ. Microbiol
- Chow, J. J., C. A. Batt, and A. J. Sinskey. 1988. **Characterization of Lactobacillus bulgaricus Bacteriophages Chapter 2**. Appl. Environ. Microbiol. 54:1138-1142
- Deri, A. 2008. **Jenis/Macam Daur Infeksi Virus (Litik & Lisogenik) + Contoh Virus Pada Hewan dan Tumbuhan**. www.organisasi.org. Diaskes tanggal 30 Januari 2012

- Djaafar, F. dan S. Rahayu. 2004. **Cemaran Mikroba**. www.pustaka.litbang.deptan.go.id. Diakses tanggal 30 Januari 2012.
- Ellis, B. 2007. **Isolation Potential MS2 Bacteriophage Strains**. www.graduate.ua.edu. Diakses tanggal 11 Februari 2012
- Garcia, P, B. Martinez, J. Obeso, A. Rodriguez. 2008. **Bacteriophages And Their Application In Food Safety**. www.cat.inist.fr. Diakses tanggal 30 Januari 2012
- Greer, G. G., and B. D. Dilts. 2002. **Control of *Brochothrix thermosphacta* Spoilage of Pork Adipose Tissue Using Bacteriophages**. J. Food Prot. 65:861-863
- Gorski, A., R. Miedzybrodzki, J. Borysowski, B. Weber-Dabrowska, M. Lobočka, W. Fortuna, S. Letkiewicz, M. Zimecki, G. Filby. 2009. **Bacteriophage Therapy For The Treatment Of Infections**. Current Opinion in Investigational Drugs (London, England : 2000) 10 (8): 766-74
- Guenther, S., D. Huwyler, S. Richard, dan M. Loessner. 2008. **Virulent Bacteriophage for Efficient Biocontrol of *Listeria monocytogenes* in Ready-To-Eat Foods**. www.ebifoodsafety.com. Diakses tanggal 30 Januari 2012
- Hagens, S, and J. Martin. 2007. **Application of Bacteriophages for detection and control of foodborn pathogens**. www.ebifoodsafety.com. Diakses tanggal 29 Januari 2012
- Leverentz, B, W. Conway, W. Janisiewicz, and M. Camp. 2004. **Optimizing Concentration and Timing of a Phage Spray Application To Reduce *Listeria monocytogenes* on Honeydew Melon Tissue**. www.ucce.ucdavis.edu. Diakses 1 Mei 2011
- Lu, Z., Breidt Jr F., Fleming H., Altermann E., Klaenhammer T. 2003. **Isolation and Characterization of a *Lactobacillus plantarum* Bacteriophage, φ JL-1, From Cucumber Fermentation**. www.sciencedirect.com. Dilihat tanggal 21 September 2011
- Mayer, G. 2010. **Bacteriology Chapter Seven : Bacteriophage, The Board of Trustees of the University of South Carolina**. www.pathmicro.med.sc.edu.
- Modi R, Y. Hirvi, A. Hill, M. Griffiths. 2001. **Effect of phage on survival of *Salmonella enteritidis* during manufacture and storage of cheddar cheese made from raw and pasteurized milk**. www.j.foods.com. Diakses 1 Mei 2011
- Nainggolan, JF. 2009. **Masalah Kesehatan Akibat Foodborn Disease**. www.mdopost.com. Diakses tanggal 29 Januari 2012
- Nugroho, W.S. 2005. **Tingkat cemaran *Salmonella* sp. pada telur ayam ras di tingkat peternakan Kabupaten Sleman Yogyakarta. Prosiding Lokakarya Nasional Keamanan Pangan Produk Peternakan, Bogor, 14 September 2005**. Pusat Penelitian dan Pengembangan Peternakan, Bogor. hlm. 160–165.
- NSW Health. 2008. **Foodborne Disease**. www.health.nsw.gov.au. Diakses tanggal 29 Januari 2012

- NSW Health. 2008. **Salmonellosis**. www.health.nsw.gov.au. Diakses tanggal 29 Januari 2012
- NSW Health. 2010. **Salmonella**. www.health.nsw.gov.au. Diakses tanggal 10 Februari 2012
- Raquel, M, and F. Yokoya. 2004. **Lactococcus Bacteriophages Isolated from Whey and Their Effects on Comercial Lactic Starters**. www.scielo.br. Diakses tanggal 29 Januari 2012
- Sharma, Manan. 2008. **Bacteriophage as Novel Antimicrobials for Food Safety**. USDA-ARS Food Safety Laboratory. www.ars.usda.gov.
- Sulakvelidze, Alexander, Zemphira, Alavidze, and J. Glenn Morris Jr. 2001. **Bacteriophage Therapy**. www.aac.asm.org. Diakses 30 Januari 2012.
- United States Food and Drug Administration. 2006. **Agency Response Letter GRAS Notice No. GRN 000198**. www.fda.gov. Diakses tanggal 29 Januari 2012
- Watson, J.D., N.H. Hopkins, J.W. Roberts, J.A Steitz, A.M. Weiner. 1987. **Molecular Biology of the Gene**. Menlo Park: Benjamin Cummings Company, Inc., CA
- Whichard, J.M., N. Sriranganathan, F.W. Pierson. 2003. **Suppression of Salmonella growth by wild-type and large-plaque variants of bacteriophage Felix O1 in liquid culture and on chicken frankfurters**. www.ebifoodsafety.com. Diakses tanggal 30 Januari 2012
- Zein, R. 2011. **Isolasi Bakteriofag dari Sampel Usus**. Skripsi Mikrobiologi Umum. Brawijaya Malang